LYN A, Active
Full-length recombinant protein expressed in Sf9 cells

Catalog # L13-18G
Lot # L028-1

Product Description
Recombinant full-length human LYN A was expressed by baculovirus in Sf9 insect cells using an N-terminal GST tag. The gene accession number is NM_002350.

Gene Aliases
JTK8, FLJ26625

Formulation
Recombinant protein stored in 50mM Tris-HCl, pH 7.5, 150mM NaCl, 10mM glutathione, 0.1mM EDTA, 0.25mM DTT, 0.1mM PMSF, 25% glycerol.

Storage and Stability
Store product at −70°C. For optimal storage, aliquot target into smaller quantities after centrifugation and store at recommended temperature. For most favorable performance, avoid repeated handling and multiple freeze/thaw cycles.

References
LYN A is a member of the Src-family of protein kinases that participate in signaling pathways of a variety of cell surface receptors and are localized to the cytoplasmic side of the plasma membrane (1). LYN A is biosynthetically transported to the plasma membrane via the Golgi pool of caveolin along the secretory pathway. The trafficking of LYN A from the Golgi apparatus to the plasma membrane is inhibited by deletion of the kinase domain but not by kinase inactivation. The kinase domain of LYN A plays a role in LYN A trafficking besides catalysis of substrate phosphorylation. cDNA microarray analysis revealed that LYN A may be involved in the changes that are related to tumor pathogenesis (2).

References

Specific Activity
The specific activity of LYN A was determined to be 510 nmol/min/mg as per activity assay protocol.

Purity
The purity of LYN A was determined to be >95% by densitometry, approx. MW 81kDa.

LYN A, Active
Full-length recombinant protein expressed in Sf9 cells

Catalog Number L13-18G
Specific Activity 510 nmol/min/mg
Specific Lot Number L028-1

Purity
>95%
0.1 µg/µl
1yr At −70°C from date of shipment
Store product at −70°C. For optimal storage, aliquot target into smaller quantities after centrifugation and store at recommended temperature. For most favorable performance, avoid repeated handling and multiple freeze/thaw cycles. Product shipped on dry ice.

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Activity Assay Protocol

Reaction Components

**Active Kinase** (Catalog #: L13-18G)

Active LYN A (0.1µg/µl) diluted with Kinase Dilution Buffer IV (Catalog #: K24-09) and assayed as outlined in sample activity plot. (Note: these are suggested working dilutions and it is recommended that the researcher perform a serial dilution of Active LYN A for optimal results).

**Kinase Dilution Buffer IV** (Catalog #: K24-09)

Kinase Assay Buffer II (Catalog #: K02-09) diluted at a 1:4 ratio (5X dilution) with final 50ng/µl BSA solution.

**Kinase Assay Buffer II** (Catalog #: K02-09)

Buffer components: 25mM MOPS, pH 7.2, 12.5mM β-glycerol-phosphate, 20mM MgCl₂, 12.5mM MnCl₂, 5mM EGTA, 2mM EDTA. Add 0.25mM DTT to Kinase Assay Buffer prior to use.

**Substrate** (Catalog #: P61-58)

Poly (4:1 Glu, Tyr) synthetic peptide substrate diluted in distilled H₂O to a final concentration of 1mg/ml.

**[³³P]-ATP Assay Cocktail**

Prepare 250µM [³³P]-ATP Assay Cocktail in a designated radioactive working area by adding the following components: 150µl of 10mM ATP Stock Solution (Catalog #: A50-09), 100µl [³³P]-ATP (1mCi/100µl), 5.75ml of Kinase Assay Buffer II (Catalog #: K02-09). Store 1ml aliquots at -20°C.

**10mM ATP Stock Solution** (Catalog #: A50-09)

Prepare ATP stock solution by dissolving 55mg of ATP in 10ml of Kinase Assay Buffer II (Catalog #: K02-09). Store 200µl aliquots at -20°C.

**Assay Protocol**

1. Thaw [³³P]-ATP Assay Cocktail in shielded container in a designated radioactive working area.
2. Thaw the Active LYN A, Kinase Assay Buffer, Substrate and Kinase Dilution Buffer on ice.
3. In a pre-cooled microfuge tube, add the following reaction components bringing the initial reaction volume up to 20µl:
   - Component 1. 10µl of diluted Active LYN A (Catalog #L13-18G)
   - Component 2. 5µl of 1mg/ml stock solution of substrate (Catalog #P61-58)
   - Component 3. 5µl distilled H₂O (4°C)
4. Set up the blank control as outlined in step 3, excluding the addition of the substrate. Replace the substrate with an equal volume of distilled H₂O.
5. Initiate the reaction by the addition of 5 µl [³³P]-ATP Assay Cocktail bringing the final volume up to 25µl and incubate the mixture in a water bath at 30°C for 15 minutes.
6. After the 15 minute incubation period, terminate the reaction by spotting 20 µl of the reaction mixture onto individual pre-cut strips of phosphocellulose P81 paper.
7. Air dry the pre-cut P81 strip and sequentially wash in a 1% phosphoric acid solution (dilute 10ml of phosphoric acid and make a 1L solution with distilled H₂O) with constant gentle stirring. It is recommended that the strips be washed a total of 3 intervals for approximately 10 minutes each.
8. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
9. Determine the corrected cpm by removing the blank control value (see Step 4) for each sample and calculate the kinase specific activity as outlined below.

**Calculation of [P³³]-ATP Specific Activity (SA) (cpm/pmol)**

Specific activity (SA) = cpm for 5 µl [³³P]-ATP / pmoles of ATP (in 5 µl of a 250 µM ATP stock solution, i.e., 1250 pmoles)

**Kinase Specific Activity (SA) (pmol/min/µg or nmol/min/mg)**

Corrected cpm from reaction / [(SA of [³³P]-ATP in cpm/pmol) * (Reaction time in min) * (Enzyme amount in µg or mg)] / [(Reaction Volume) / (Spot Volume)]

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